

TRIIODOTHYRONINE STIMULATES 3 β -HYDROXYSTEROID DEHYDROGENASE ACTIVITY IN THE PORCINE CORPUS LUTEUM

E.L. GREGORASZCZUK, J. KOŁODZIEJCZYK, ¹J. RZYSA

Department of Animal Physiology, Institute of Zoology, Jagiellonian University, 30-060 Krakow, Poland and

¹Department of Animal Physiology, Academy of Agriculture, Kraków, Poland

Objective. To study the mechanism of thyroid hormone action on the activity of 3 β -hydroxysteroid dehydrogenase in the porcine corpus luteum.

Methods. Pig ovaries were obtained from slaughterhouse animals. Luteal cells were isolated from mid-developing (5-7 days after ovulation) corpora lutea and incubated for 24 h with or without triiodothyronine. Trilostane, an inhibitor of 3 β -HSD that blocks the conversion of pregnenolone to progesterone, was added to the medium in doses of 0, 0.1, 1, 10, and 100 μ mol. Each treatment was performed in triplicate and each culture system was set up in triplicate. Progesterone concentrations in culture media were determined by radioimmunoassays.

Results. Trilostane in a dose of 100 μ M significantly decreased the basal progesterone secretion from luteal cells by 26 % ($P < 0.05$). However, such secretion was increased by triiodothyronine (T_3) in a dose of 10^{-9} M. In addition, in T_3 -treated cells dose dependent inhibitory effect of trilostane on progesterone secretion was observed. Control cultures grown in control medium revealed a relatively weak 3 β -HSD activity which, however, markedly increased after the addition of T_3 to the culture medium. Trilostane remarkably decreased 3 β -HSD activity in T_3 -stimulated cells.

Conclusion. It was found that T_3 acts on luteal cell steroidogenesis via the activation of 3 β -hydroxysteroid dehydrogenase in these cells.

Keys words: Triiodothyronine – Corpus luteum – Progesterone – 3 β -hydroxysteroid dehydrogenase – Trilostane

Thyroid hormones are known to exert diverse effects on growth, development and metabolism of nearly all tissues. Possible relationships between thyroid hormones and ovarian steroidogenesis have been repeatedly reported (GOLDMAN et al. 1993; GREGORASZCZUK and SKALKA 1996; GREGORASZCZUK 1996; GREGORASZCZUK and GALAS 1998). Triiodothyronine (T_3) binding sites have been found in the nuclei of granulosa (GOLDMAN et al. 1993; WAKIM et al. 1994a,b) and luteal cells (BHATTACHARYA et al. 1988). We have recently shown (GREGORASZCZUK 1996) an increased progesterone production by porcine luteal cells isolated from early-developing and mid-developing corpora lutea when triiodothyronine was added to the culture medium. DATTA et al. (1998) showed that the decrease of thyroid hormone level during

the luteal phase of the menstrual cycle (between days 18 and 23 of the cycle) in women is related to a decrease in progesterone secretion.

The enzyme 3 β -hydroxysteroid dehydrogenase-isomerase (3 β -HSD) converts 5-ene-3 β -hydroxysteroids to the 4-ene-3-oxo configuration and therefore plays an essential role in the biosynthesis of hormonally active steroids such as progesterone. Progesterone synthesis by corpus luteum is a complex process involving the uptake, storage and utilisation of cholesterol derived primarily from circulating low-density lipoproteins (SANDERS and STOUFFER 1995). The initial enzymatic step in the biosynthesis of progesterone involves the conversion of cholesterol to pregnenolone by cytochrome P450_{scc}. Our previous findings have shown that cytochrome P450_{scc}

may be a key target of T₃ action on the mitochondria of luteal cells (GREGORASZCZUK and PIEKLO 1998). Then, pregnenolone is converted to progesterone by 3 β -hydroxysteroid dehydrogenase (STRAUSS III and MILLER 1991).

In porcine granulosa cells *in vitro*, thyroid hormone has been shown to enhance FSH action on aromatase (MARUO et al. 1992; GREGORASZCZUK and SKALKA 1996; GREGORASZCZUK et al. 1998). Earlier work showed the presence of thyroid hormone binding sites in the human corpus luteum (BHATACHARYA et al. 1988) and an increase in progesterone production after the addition of thyroid hormone to the porcine luteal cell culture (GREGORASZCZUK 1996). These previous studies pointed out to possible multiple sites of action of thyroid hormone on the steroidogenesis.

The current studies were initiated to determine to what extent T₃ regulates 3 β -HSD, an enzyme involved in progesterone secretion. For this purpose the classical steroidal competitive inhibitor of 3 β -HSD trilostane was tested for its effects on basal and T₃ stimulated progesterone biosynthesis.

Materials and Methods

Reagents. Triiodothyronine (10⁻⁹ M) was purchased from Sigma Chemical Co (St. Louis, MO, USA). Trilostane (4 α ,5 α ,17 β)-4,5-epoxy-3,17-hydroxyandrost-2-ene-2-carbonitrile was purchased from Sanofi Pharmaceuticals (Malvern, PA, USA). Medium M199, penicillin, trypsin, and calf serum were from the Laboratory of Vaccines (Lublin, Poland).

Animals and cell isolation. Ovaries were obtained from Large White sows from a local slaughterhouse immediately after slaughter, placed in ice-cold PBS and transported to the laboratory. The phase of the oestrous cycle was determined according to the established morphological criteria (SCHILLING 1974; GREGORASZCZUK 1992). Dissected corpora lutea from each animal were enzymatically dissociated as published previously (GREGORASZCZUK 1983). Luteal cells were obtained from pools of freshly excised mature corpora lutea (5-7 days after ovulation) from three animals to produce the luteal cell pool used in any given replicate to minimise the variation possibly existing among corpora lutea in the same ovaries and between ovaries in the same animals and among an-

imals. The cells were suspended in medium M199 supplemented with 5 % of calf serum at a concentration of 3.5x10⁵ cells/ml medium and then grown in multiwell plates (Nunk) in a humidified atmosphere with 5 % CO₂ in the air for 48 h. Incubations were conducted in triplicate in 1.0 ml/well. The cells were cultured with 10⁻⁹ M T₃. The final inhibitor concentrations used were of 0, 1, 10 and 100 μ M. After 48 h incubation, all cultures were terminated and the media were frozen until used for the steroid estimations. Cell viability measured using the trypan blue exclusion test was 85 %. At least three different experiments (n=3), each in triplicate have been performed.

Histochemical evaluation. To evaluate the influence of T₃ on the activity of delta⁵,3 β -hydroxysteroid dehydrogenase (3 β -HSD), the luteal cells were examined histochemically according to FISCHER and KAHN (1972); in this histochemical reaction the products of enzyme activity within luteal cells are formazan granules. The reaction was classified as strong (+++), intermediate (++) or weak (+) depending on the intensity of colour. The results were expressed as the percentages of cell classes with different enzyme activity. Five different areas for each treatment were measured and the number of cells was expressed as the mean \pm SEM.

Progesterone estimation. Progesterone was determined radioimmunologically using Spectria kits (Orion, Diagnostica, Finland), supplied by Polatom (Swierk, Poland). The limit of assay sensitivity was 94 pg/ml. The coefficients of variation within and between assays were 5.8 % and 2.9 %, respectively. The mean recoveries were 95.1-103.7 %. The cross-reaction with pregnenolone was 2.9 %. All other tested steroids (5 β -dihydroprogesterone, 20 β -hydroxyprogesterone, corticosterone, testosterone, estrone) showed less than 1 % cross-reaction.

Results

Progesterone secretion. The basal progesterone secretion from luteal cells was significantly decreased (P<0.05) by trilostane in a concentration of 100 μ M. In contrast, triiodothyronine in a dose of 10⁻⁹ M increased progesterone secretion from luteal cells (4.7 ng/10⁵ cells vs. 3.2 ng/10⁵ cells in the control culture). In T₃ stimulated cells a dose dependent

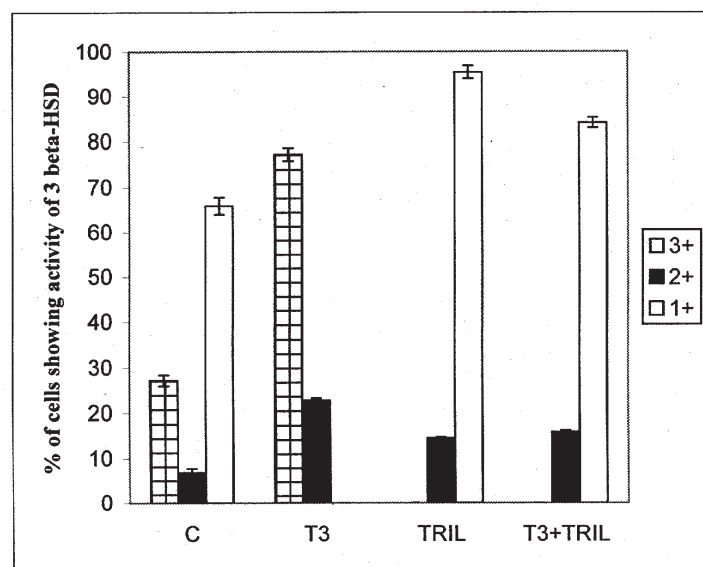


Fig. 1 The influence of trilostane (Tril) in a dose of 1, 10 or 100 μ M on basal progesterone (C) and triiodothyronine stimulated (T3) progesterone secretion by luteal cells

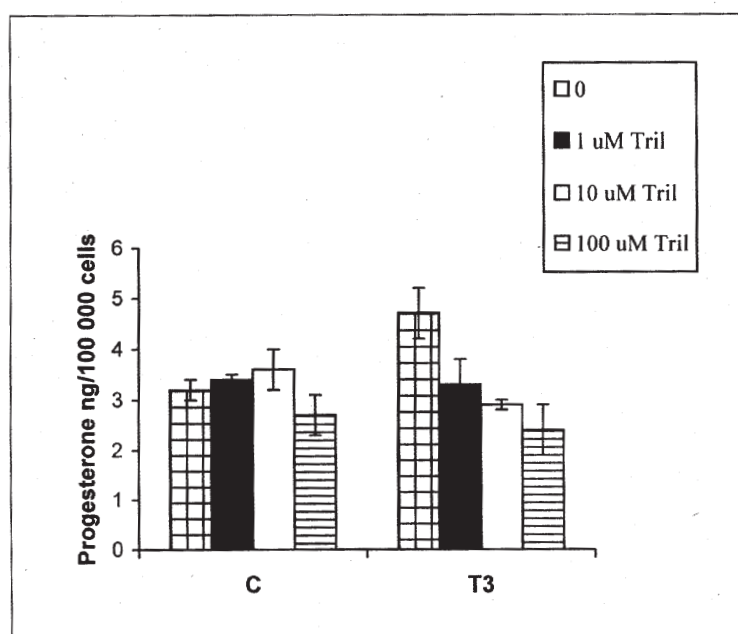


Fig. 2 The influence of T₃ on the activity of delta⁵,3 β -hydroxysteroid dehydrogenase (3 β -HSD) of luteal cells. The reaction was classified as strong (3+), intermediate (2+), or weak (1+) depending on the intensity of colour. The results were expressed as the percentage of cell classes with different enzyme activity. Five different areas for each treatment were measured and the number of cells was expressed as the mean \pm S.E.
C – control; T3 – triiodothyronine; TRIL – trilostane

inhibitory effect of trilostane on progesterone secretion was observed (30 %, 39 % and 49 % of basal secretion after trilostane concentrations of 1, 10 and 100 μ M; Fig. 1).

Histochemical results. Control cultures grown in M199 medium revealed a relatively weak enzyme activity (Fig. 2a). However, such activity markedly increased after the addition of T₃ to the culture me-

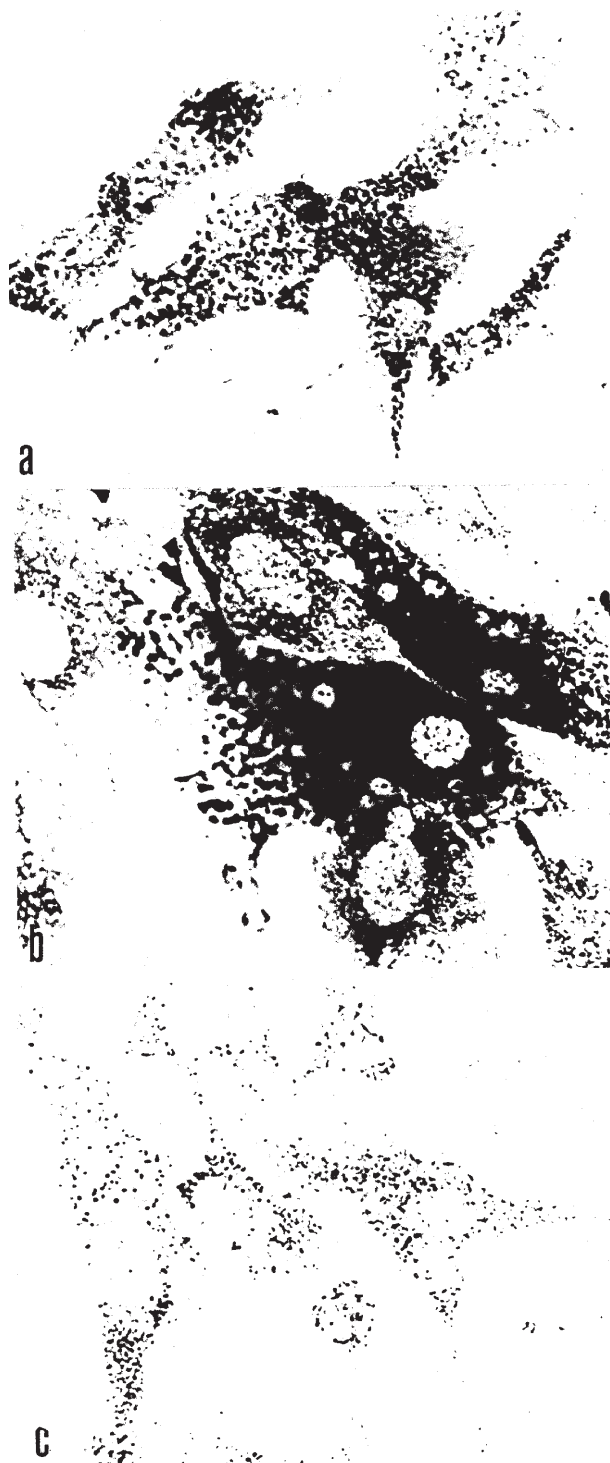


Fig. 3 Luteal cells culture tested for the delta⁵,3 β -hydroxysteroid dehydrogenase activity.

a – control cells showing weak histochemical reaction, x 320;
 b – corresponding culture in medium with addition of triiodothyronine (note strong activity of dehydrogenase), x 320; c – cells cultured in medium with the addition of triiodothyronine+trilostane (decrease of the enzyme activity is visible), x 320

dium (Fig. 2b). Trilostane, the well known inhibitor of 3 β -HSD, at a concentration of 100 μ M remarkably decreased enzyme activity (Fig. 2c) and, in addition, T₃-stimulated activity of 3 β -HSD was inhibited by all doses of trilostane used (Fig.2d).

Discussion

Progesterone synthesis by corpus luteum is a complex process involving the uptake, storage, and utilisation of cholesterol. The initial enzymatic step in the biosynthesis of progesterone involves the conversion of cholesterol to pregnenolone by cytochrome P450_{SCC} (SANDERS and STOUFFER 1995). Pregnenolone is then converted to progesterone by 3 β -hydroxysteroid dehydrogenase (STRAUSS III and MILLER 1991). Previous studies (GREGORASZCZUK and PIEKLO 1998) have shown that cytochrome P450_{SCC} may be a key target of T₃ action on the mitochondria in luteal cells. On the basis of the current study we hypothesise that 3 β -hydroxysteroid dehydrogenase-isomerase plays also an essential role in the triiodothyronine action on luteal cells. Actually, triiodothyronine induced an increase both in progesterone secretion and 3 β -HSD activity in cultures of luteal cells. This is in agreement with the data obtained by BHATTACHARYA et al. (1996) who showed that the addition of T₃ to the fish follicle incubation caused a two-fold increase of ³H-pregnenolone conversion to radiolabelled progesterone as compared to the control. SIMONIAN (1986) showed an interaction between ACTH and thyroid hormone for the stimulation of 3 β -HSD activity in human fetal adrenal cortex cultured cells. Trilostane have been shown to be a competitive inhibitor of 3 β -HSD by several different sources (NAVILLE et al. 1991; COOKE 1996). The studies presented here demonstrate that in the porcine corpus luteum, this compound also acts as an competitive inhibitor of the 3 β -HSD mediated step in progesterone biosynthesis.

The present study demonstrates that trilostane essentially abolished basal progesterone secretion and 3 β -HSD activity in cultured luteal cells. HAYASHI et al. (1987) using a monolayer culture system of porcine granulosa cells obtained from small porcine follicles showed that thyroid hormone increases FSH-mediated induction of 3 β -hydroxysteroid dehydrogenase in immature granulosa cells. Recently pub-

lished observations by DATTA et al. (1999) indicated that the addition of increasing concentrations of T₃ caused a linear increase of 3 β -HSD activity.

Steroidogenesis is controlled by a number of factors, the most significant of which are availability of the substrate (MURPHY and SILAVIN 1989), and the effect of the cAMP cascade on cholesterol side chain cleavage (HALL 1985). However, there is good evidence that trophic regulation of steroid synthesis may occur also by the alteration of the activity of appropriate enzymes, including 3 β -HSD. Administration of T₃ to the culture medium of luteal cells stimulated 3 β -HSD activity over untreated control. Trilostane added together with triiodothyronine reduced progesterone secretion and 3 β -HSD activity when compared to cultures treated with triiodothyronine alone.

In summary, the present study demonstrated that T₃ influences steroidogenesis in the corpus luteum by stimulating 3 β -HSD activity and this regulation is a component of the luteotrophic regulation of steroidogenesis in the luteal cells. Effective inactivation of 3 β -HSD prevents the stimulatory effect of T₃ on progesterone secretion from luteal cells. Further studies are necessary to show if the effect of T₃ on luteal progesterone secretion is direct or, as it shown by BHATTACHARYA et al. (1996) for perch ovarian follicles, by the induction of generation of a novel putative protein in ovarian follicles that stimulates the conversion of pregnenolone to progesterone.

Acknowledgements

The authors wish to thank Professor Z. Dobrowski for his constant interest and Dr.M.Mika for radioimmunological determinations of steroid hormones. This work was supported by the State Committee for Scientific Research (KBN) grant no 5P06D 030 14.

References

- BHATTACHARYA J, DANERJEE J, JAMALUDDIN M, BANERJEE PP, MAITRA G: Thyroid hormone binding to human corpus luteum. *Experientia* **44**, 1005-1007, 1988
- BHATTACHARYA S, GUIN S, BANDYOPADHYAY A, JANA NR, HALDER S: Thyroid hormone induces the generation of a novel putative protein in piscine ovarian follicle that stimulates the conversion of pregnenolone to progesterone. *Eur J Endocrinol* **134**, 128-35, 1996
- COOKE GM: Differential effect of trilostane and cyanoketone on the 3 β -hydroxysteroid dehydrogenase-isomerase reactions in androgen and 16-androstene biosynthetic pathways in the pig testis. *J Steroid Biochem Molec Biol* **58**, 95-101, 1996
- DATTA M, NAGENDA PRASAD RJ, BHATTACHARYA S: Thyroid hormone regulation of perch ovarian 3 β -hydroxysteroid dehydrogenase/delta5-delta4-isomerase activity: involvement of 52-kDa protein. *Gen Comp Endocrinol* **113**, 212-20, 1999
- DATTA M, ROY P, BANERJEE J, BHATTACHARYA S: Thyroid hormone stimulates progesterone release from human luteal cells by generating a proteinaceous factor. *J Endocrinol* **158**, 319-25, 1998
- FISCHER TV, KHAN RH: Histochemical studies of rat ovarian follicular cells in vitro. *In Vitro* **7**, 201-205, 1972
- GOLDMAN S, DIRNFELD M, ABRAMOVICI H, KRAIEM Z: Triiodothyronine (T₃) modulates hCG-regulated progesterone secretion, cAMP accumulation and DNA content in cultured human luteinized granulosa cells. *Mol Cell Endocr* **96**, 125-131, 1993
- GREGORASZCZUK EL, GALAS J: Triiodothyronine-dependent activation of cAMP accumulation and steroids secretion by follicular and thecal cells. *Endocrine Regul* **32**, 93-98, 1998
- GREGORASZCZUK EL, PIEKLO R: Thyroid hormone action in porcine luteal cells. Effect of triiodothyronine on mitochondrial cytochrome P450-SCC activity. *J Physiol Pharmacol* **49**, 467-475, 1998
- GREGORASZCZUK EL, SKALKA M: Thyroid hormone as a regulator of basal and human chorionic gonadotrophin-stimulated steroidogenesis by cultured porcine theca and granulosa cells isolated at different stages of the follicular phase. *Reprod Fertil Dev* **8**, 961-967, 1996
- GREGORASZCZUK EL, SŁOMCZYŃSKA M, WILK R: Thyroid hormone inhibits aromatase activity in porcine thecal cells cultured alone and in coculture with granulosa cells. *Thyroid* **8**, 1157-1163, 1998
- GREGORASZCZUK EL: Interrelationship between steroid hormone secretion and morphological changes of porcine corpora lutea at various periods of luteal phase. *Endocrine Regul* **26**, 189-194, 1992
- GREGORASZCZUK EL: Steroid hormone release in cultures of pig corpus luteum cells. Effect of LH, hCG, PRL and estradiol. *Endocrinol Exp* **17**, 59-63, 1983
- GREGORASZCZUK EL: The effect of triiodothyronine (T₃) and TSH treatment in vitro on progesterone production by luteal cells isolated during different stages of the luteal phase. *J Physiol Pharmacol* **47**, 93-100, 1996

- HALL PF: Trophic stimulation of steroidogenesis: In search of the elusive trigger. *Rec Prog Horm Res* **41**:1, 1985
- HAYASHI M, MARUO T, MATSUO H, MOCHIZUKI M: Effect of thyroid hormone on steroidogenic enzyme induction in porcine granulosa cells cultured in vitro. *Nippon Naibunpi Gakkai Zasshi* **63**, 1231-40, 1987
- MARUO T, HIRAMATSU S, OTANI T, HAYASHI M, MOCHIZUKI M: Increase in the expression of thyroid hormone receptors in porcine granulosa cells early in follicular maturation. *Acta Endocrinol* **127**, 152-160, 1992
- MURPHY BD, SILAVIN S: Luteotrophic agents and steroid substrate utilization. *Oxford Rev Reprod Biol* **11**, 180, 1989
- NAVILLE D, KEENEY DS, JENKIN G, MURRY BA, HEAD JR, MASON JI: Regulation of expression of male-specific rat liver 3 β -hydroxysteroid dehydrogenase. *Mol Endocrinol* **5**, 1090-1100, 1991
- SANDERS SL, STOUFFER RL: Gonadotropin- and lipoprotein-supported progesterone production by primate luteal cell types in culture. *Endocrine* **3**, 169-175, 1995
- SCHILLING E: Stages of ovarian function in the sow. In: *Veterinary Medical Review* (Elwert EG, ed.), pp. 59-63, University und Verlagsbuchhandlung, Marburg/Lahn 1974
- SIMONIAN MH: ACTH and thyroid hormone regulation of 3 β -hydroxysteroid dehydrogenase activity in human fetal adrenocortical cells. *J Steroid Biochem* **25**, 1001-6, 1986
- STRAUSS III JF, MILLER WL: Molecular basis of ovarian steroid synthesis. In: Hillier SG (ed.), *Ovaria Endocrinology*, pp.25-72, Blackwell Scientific, Oxford 25-72, 1991
- WAKIM AN, PALJUG WR, JASNOSZ KM, ALKAHIM N, BROWN AB, BURHOLD R: Thyroid hormone receptors messenger ribonucleic acid in human granulosa and ovarian stromal cells. *Fertil Steril* **62**, 531-534, 1994a
- WAKIM AN, POLIZOTO SA, BURHOLD DR: α -1 and β -1 thyroid hormone receptors on human granulosa cells. *Rec Progr Horm Res* **49**, 377-381, 1994b
- Corresponding author:** E.L. Gregoraszczuk
Department of Animal Physiology
Institute of Zoology
Jagiellonian University
Ingardena 6, 30-060 Krakow, Poland
e-mail: greg@zukunft.iz.uj.edu.pl

Accepted: June 15, 1999